

**Bijlage 1.****Proces & tijdslijn Covid-19 vaccin-registratie en uitlevering t.b.v. gesprek met minister.**

De eerste vaccinproducenten gaan hun dossiers indienen voor markttoelating in de EU. Hieronder wordt geschetst hoe dit registratieproces en het leveringsproces de komende weken mogelijk in de tijd gaat verlopen. In dit overzicht richten we ons vooral op de vaccins van BionTech/Pfizer en Moderna. Ook zal naar verwachting Johnson&Johnson deze week zijn dossier indienen voor goedkeuring. Naar verwachting zal het registratieverzoek van AztraZeneca door omstandigheden later volgen. De verzoeken tot markttoelating vanuit CureVac en Sanofi/GSK, worden sowieso niet dit jaar verwacht. Over de indiening door J&J en de planning, is op dit moment nog geen verdere informatie voorhanden

- **Week 49 (v.a. 30/11)**

- Moderna, BionTech/Pfizer en Johnson&Johnson dienen hun dossiers voor goedkeuring in bij EMA.

- **Week 50 (v.a. 7/12)**

- **Week 51 (v.a. 14/12)**

- 15/12 mogelijke start OMCL (Official Medicines Control Laboratory)-procedure voor **Moderna**. Voor **BionTech** is deze datum nog niet bekend.
- Afronding OMCL-procedure + 10 dagen.  
*(N.b.: beide producenten vragen mogelijk een OMCL-waiver aan, waardoor die procedure, de levering nu niet hoeft te vertragen.)*
- Verzoek van mogelijk beide producenten om tijdelijke vrijstelling van FMD-procedure gericht op vervalsingen.

- **Week 52 (v.a. 21/12)**

- **Week 53 (v.a. 28/12)**

- Verwachte goedkeuring door EMA van vaccin **BionTech/Pfizer** (29/12)
- Markttoelatingsbesluit door Europese Commissie + 2 dagen.

- **Week 1 (v.a. 4/1)**

- Start vaccinatie **BionTech/Pfizer**.

- **Week 2 (v.a. 11/1)**

- Verdere uitrol vaccinatie **BionTech Pfizer**
- Verwachte goedkeuring door EMA van vaccin **Moderna** (12/1)
- Markttoelatingsbesluit **Moderna** door Europese Commissie + 2 dagen

- **Week 3 (v.a. 18/1)**

- Verdere uitrol vaccins **BionTech Pfizer**
- Start vaccinatie **Moderna**

<b>Start verwachte levering BionTech/Pfizer (op basis bestelling):</b> <b>Q4 2020: 1.050.434</b> <b>Q1 2021: 1.680.694</b>  <b>Start verwachte levering Moderna (gebaseerd op eerste indicaties):</b> <b>Q1 2021: 400.000</b>  <b>Start verwachte levering Johnson&amp;Johnson</b> <b>Q2 2021: 3.000.000</b>
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Methods Assessment for SARS-CoV-2 Genetic Signal in Wastewater:  
**WRF 5089 Study Results**

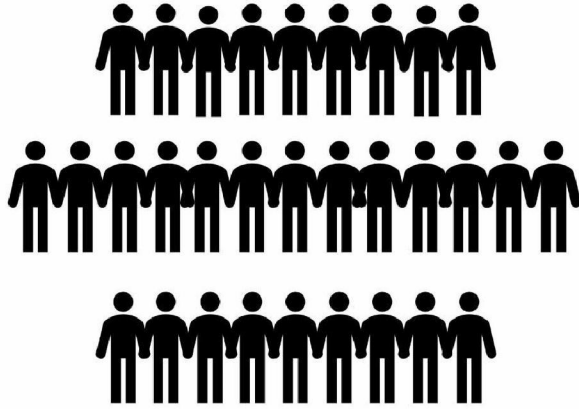


**Ph.D., P.E. – Trussell Technologies**

**P.E. – Trussell Technologies**

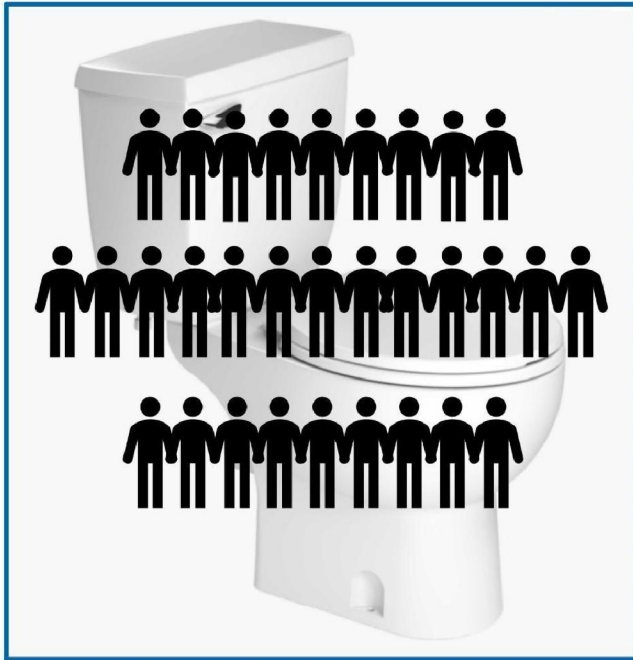
**Ph.D. – Drexel University**

# Community Health



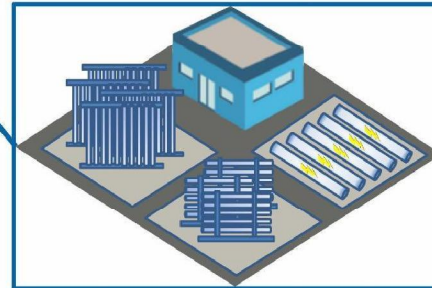
Clinical testing of individuals

# Community Health



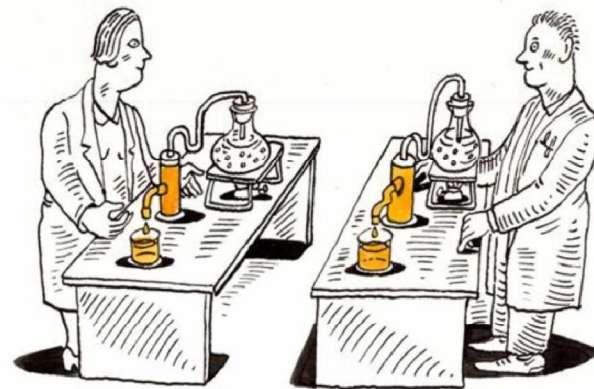
Clinical testing of individuals

Wastewater Based Epidemiology

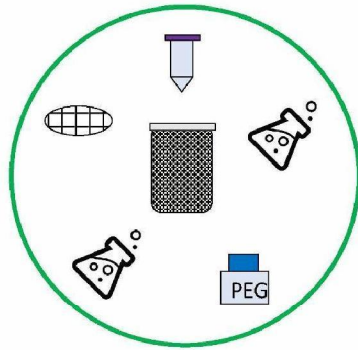


## Project Objective


- Assess the methods currently used by laboratories to determine which methods provide a reliable and repeatable measurement of the SAR-CoV-2 genetic signal in untreated wastewater



# Overview of Project



60+ labs

 **THE Water Research FOUNDATION**

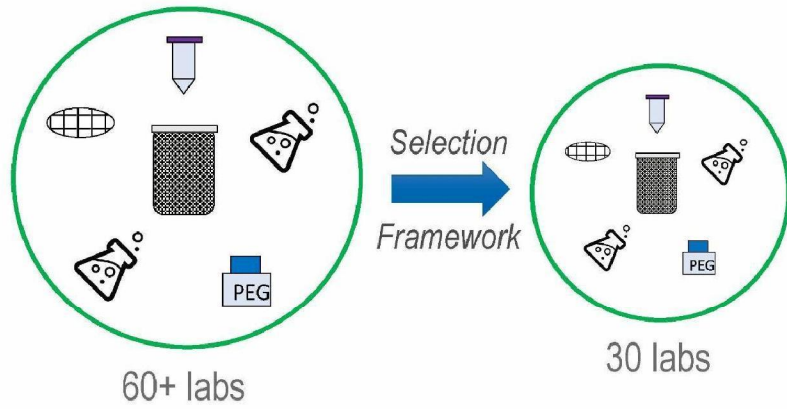
**PARTICIPATION FORM**  
**Interlaboratory and Methods Assessment of SARS-CoV-2 Genetic Signal in Wastewater (5089)**  
**Participation Form Due by 4:00 PM MDT on June 7, 2020**

First Name, Last Name: \_\_\_\_\_ Title: \_\_\_\_\_  
Email: \_\_\_\_\_ Phone: \_\_\_\_\_  
Lab/Organization Name: \_\_\_\_\_  
Street Address: \_\_\_\_\_  
City: \_\_\_\_\_ State: \_\_\_\_\_ Zip Code: \_\_\_\_\_ Country: \_\_\_\_\_

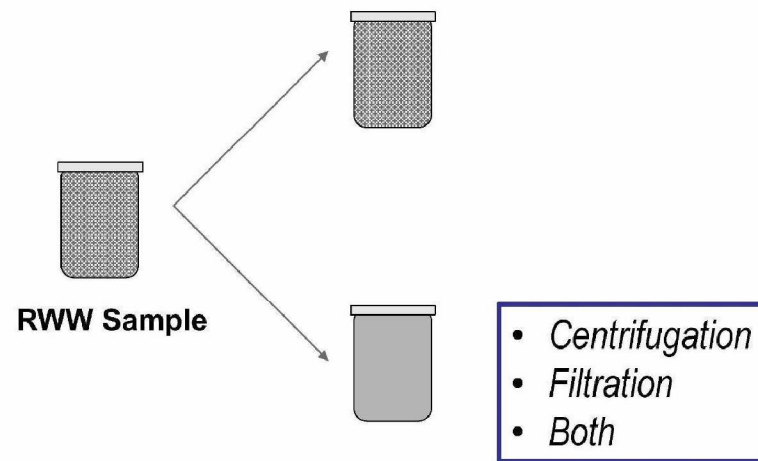
**Please verify that you meet the following minimum participating requirements:**

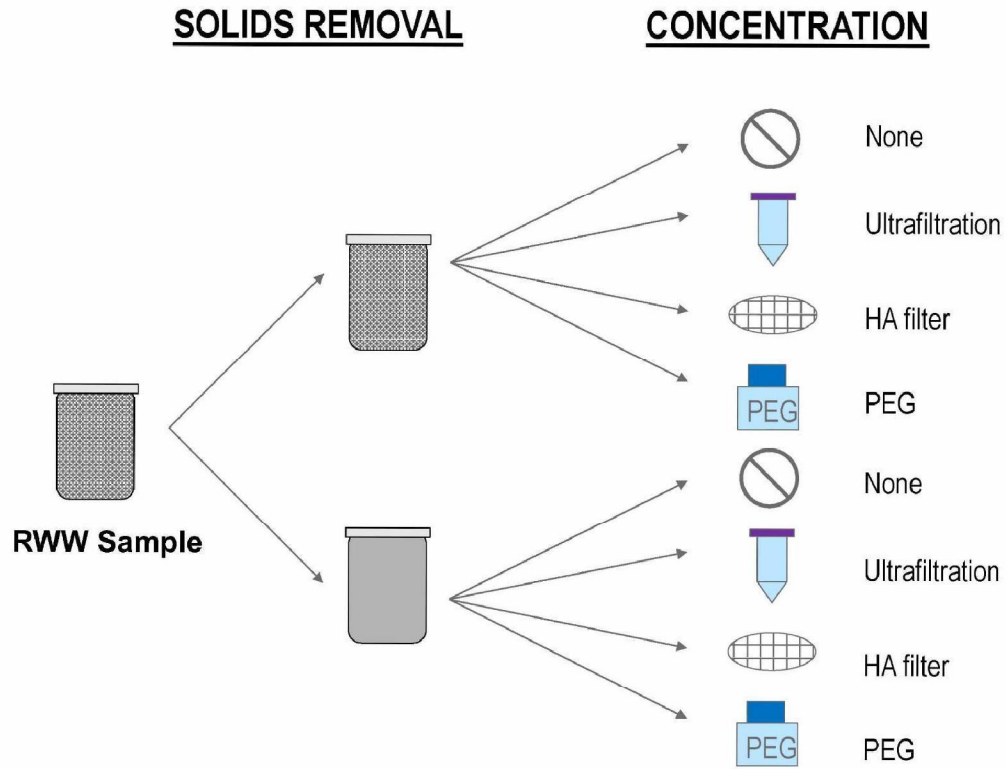
- Ability to test 5-10 samples at **their own expense**  Y  N
- Has a developed method for the detection of the genetic signal of SARS-CoV-2 (please provide written protocol and describe controls in box below) \_\_\_\_\_
- Is routinely or planning to routinely analyze samples for the genetic signal of SARS-CoV-2 for environmental surveillance  Y  N
- Ability to handle wastewater samples that have been pre-treated to inactivate live microorganisms (samples that have undergone pre-treatment (i.e., pasteurization))  Y  N
- Has the reagents and equipment to quickly process samples supplied by the selected research team  Y  N
- Is established as an environmental microbiology or research laboratory  Y  N (please provide details of accreditation in box below) \_\_\_\_\_
- Has a quality assurance plan for the overall operation of the lab that can be submitted to requesting RPO respondents  Y  N
- Has the ability to share data with the selected research team  Y  N
- In the box below, please list methods used (e.g., RT-PCR, dd-PCR, Metagenomics), including an outline of concentration, extraction, and assay \_\_\_\_\_
- Notice of any patents or proprietary equipment or methods that may be used for this project \_\_\_\_\_
- Estimated earliest start date that samples can be received for analysis: \_\_\_\_\_

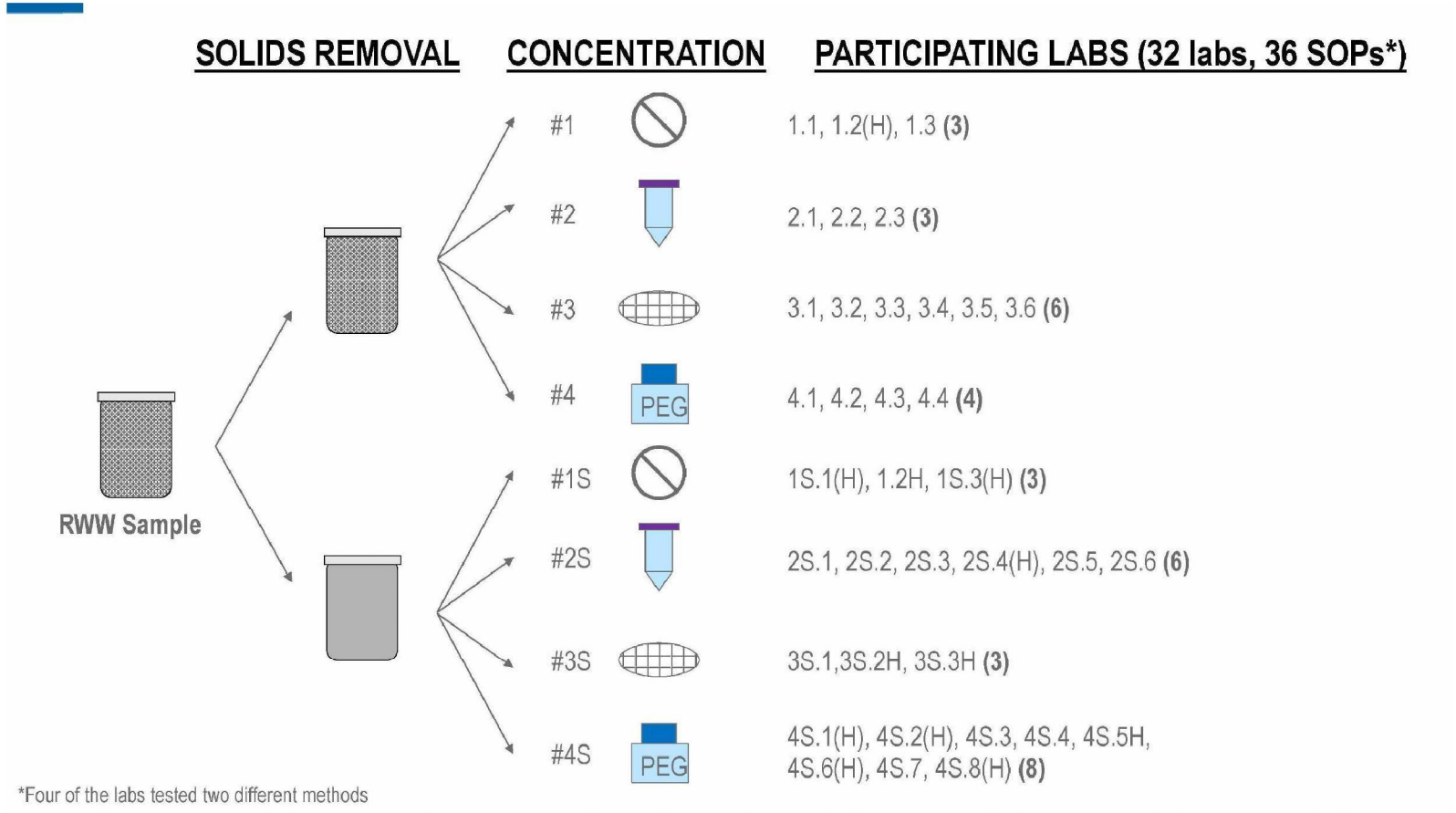
# Overview of Project



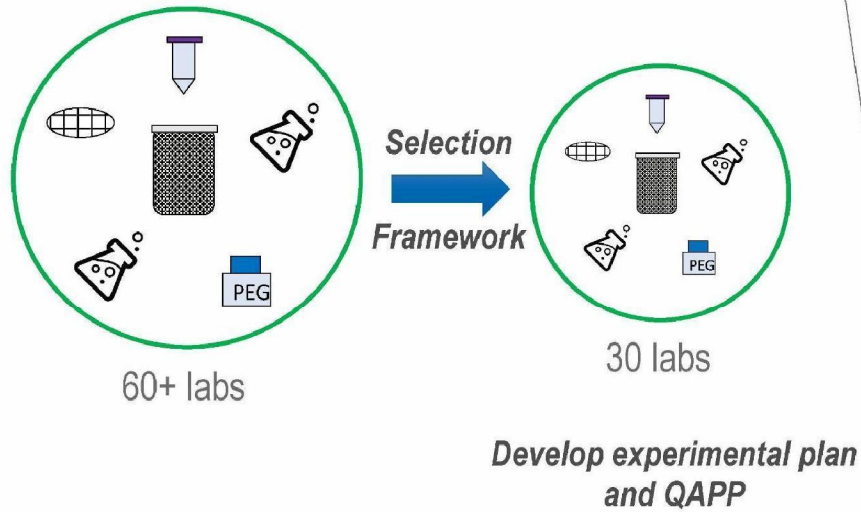
## SOLIDS REMOVAL







# Overview of Project



**Quality Assurance Project Plan**  
Analytical Microbiology Services  
Water Research Foundation  
Contract #5089  
Interlaboratory and Methods Assessment of  
Wastewater

Prepared for:  
The Water Research Foundation

Prepared by:  
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July 2020  
Version 1.0

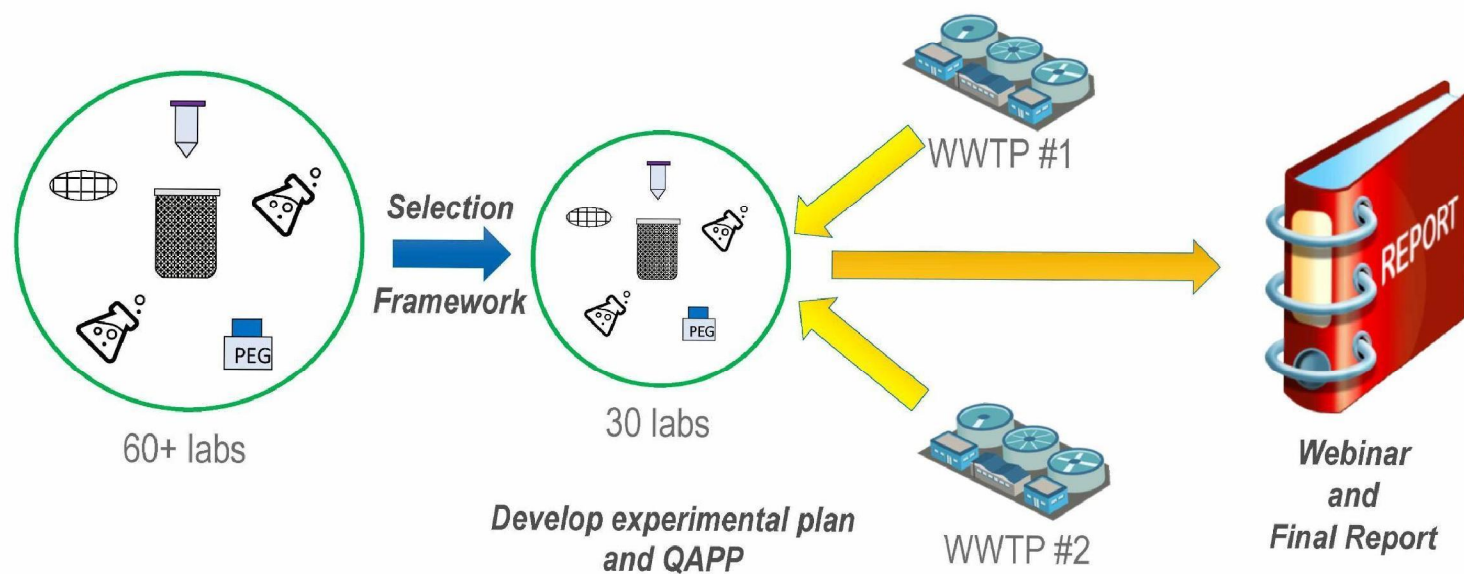
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## Wastewater Sampling



- 5 replicates per round
- 2 wastewater treatment plants
  - *Hyperion Water Reclamation Plan*
  - *Joint Water Pollution Control Plant*
- Follow sampling/shipping requirements from QAPP

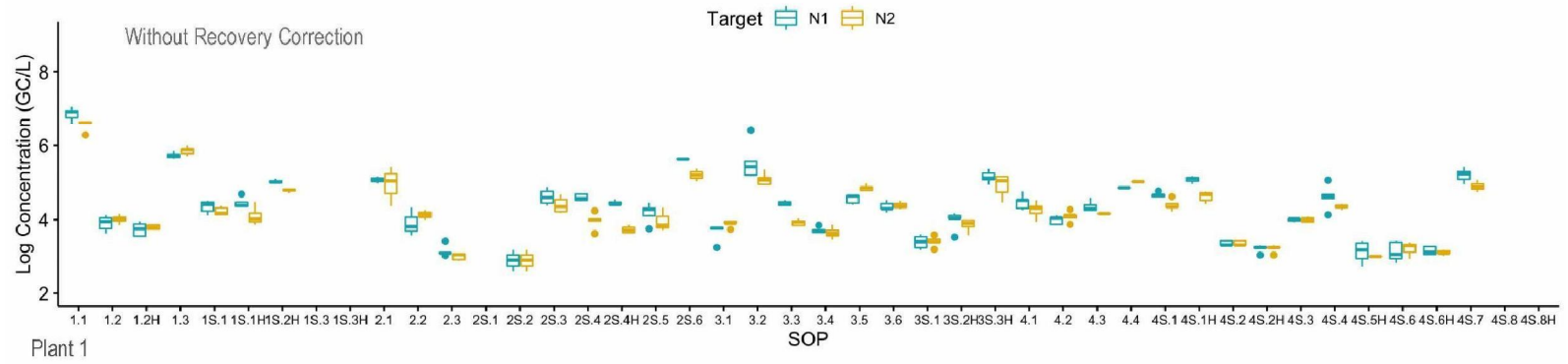
## Overview of Project





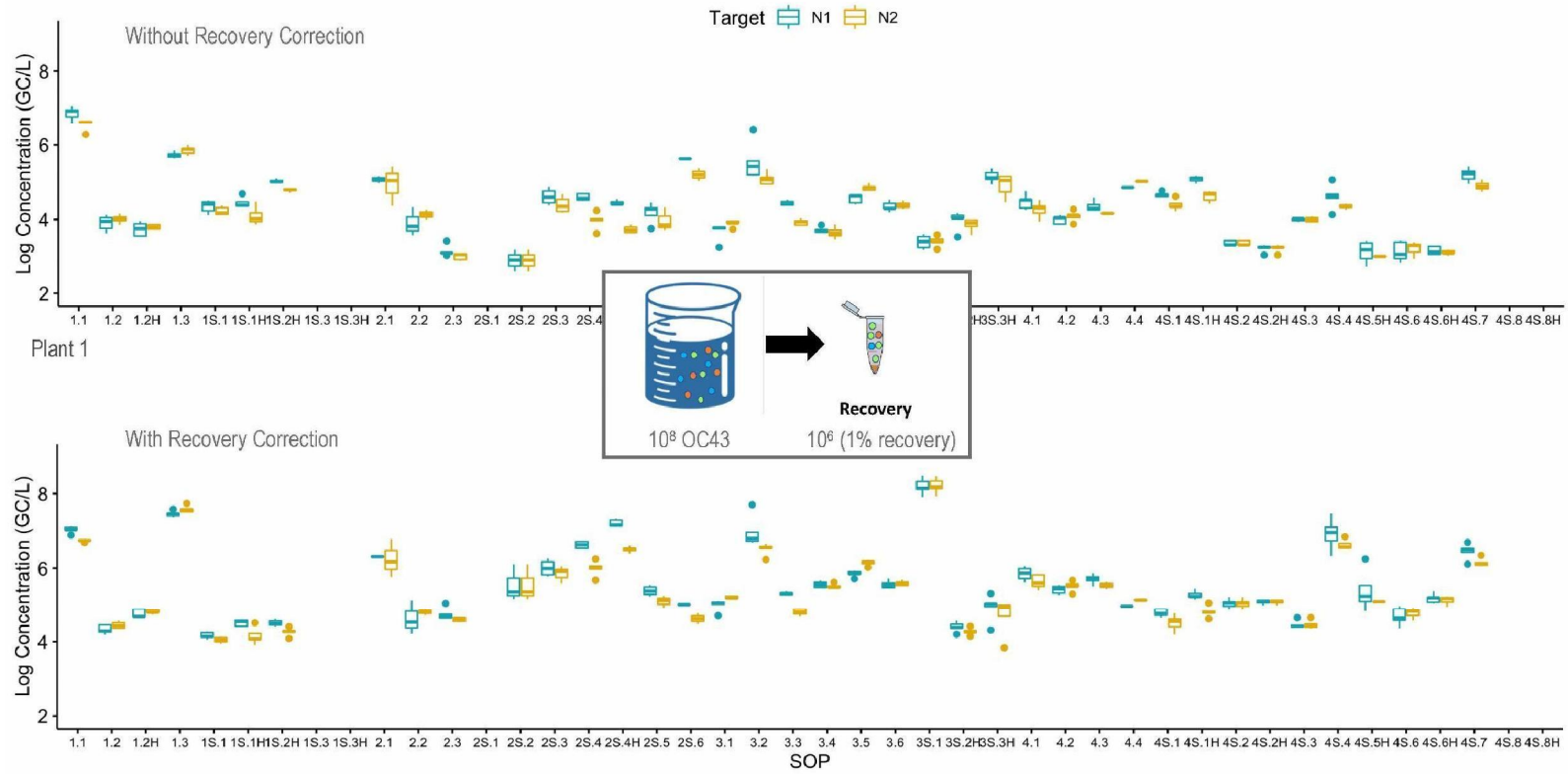
Reproducibility

# Reproducibility across methods





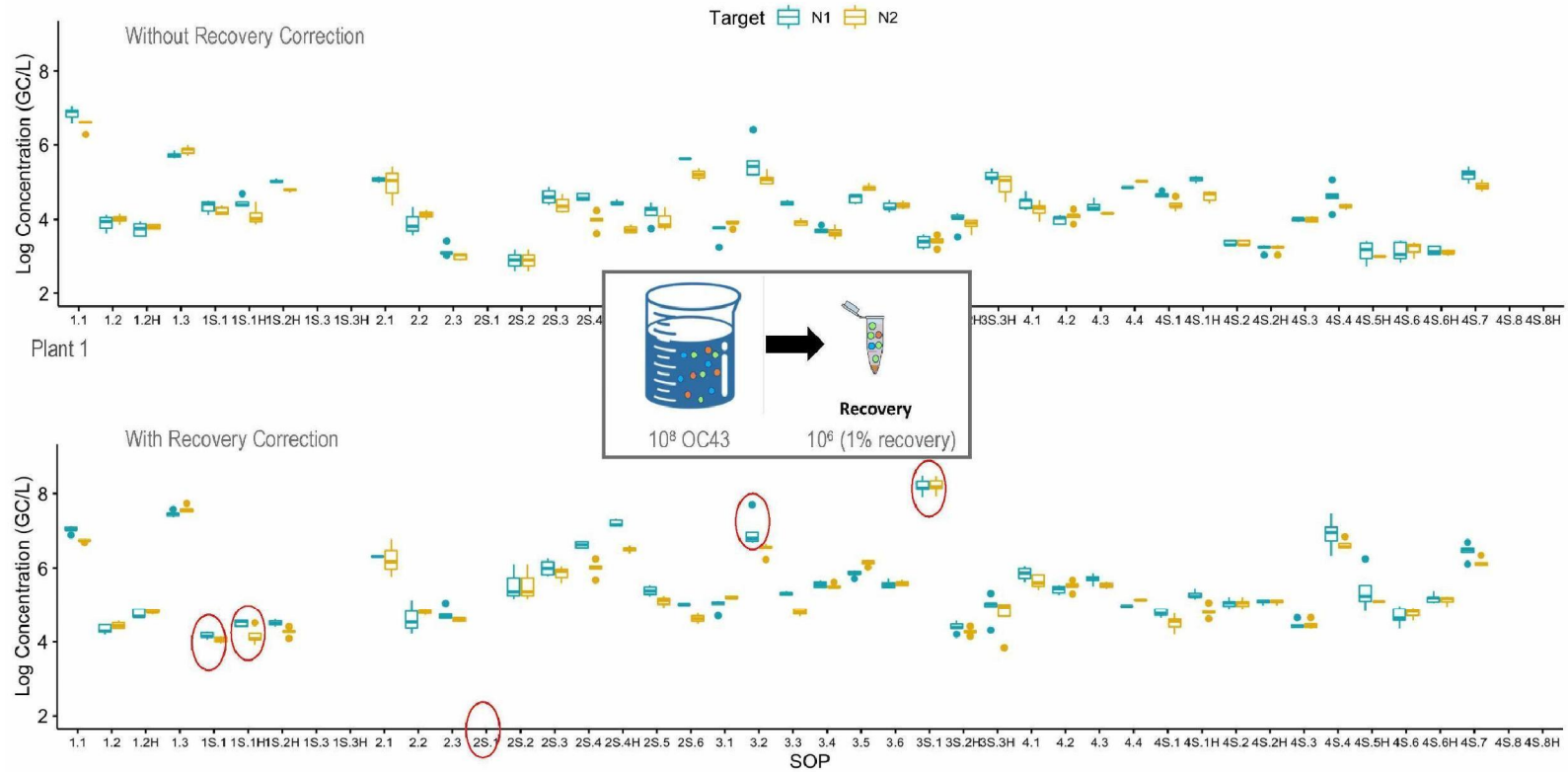
# Reproducibility across methods



## QA/QC and Exclusion Criteria

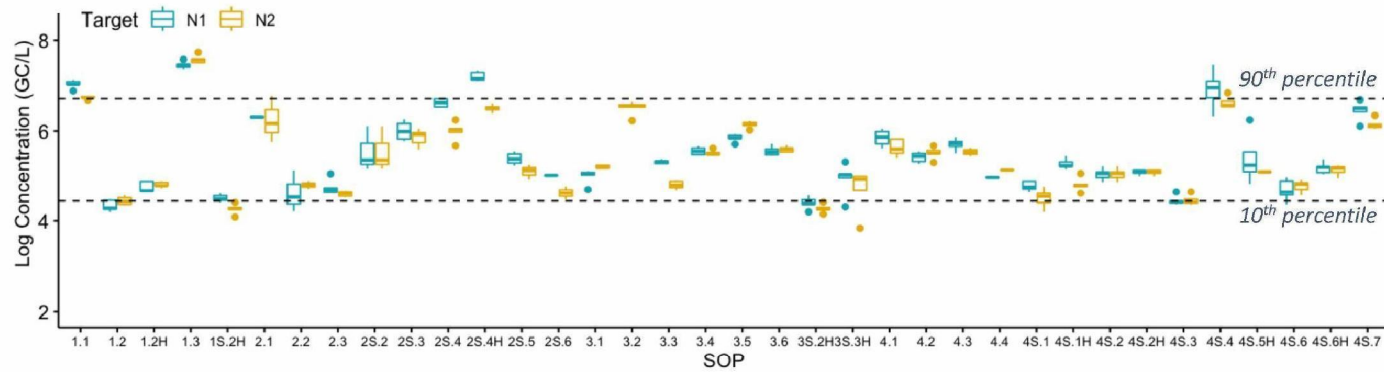
- Sample processing
  - *More than 24 h after receipt of sample*
- No-template controls
  - *All NTC replicates positive*
  - *Similar order of magnitude as environmental samples*
- Recovery efficiency
  - *Recoveries < 0.01% were rejected*
- Detection limit
  - *Results lower than the lowest detectable standard (by a factor greater than 2)*

# Reproducibility across methods



# Reproducibility across methods after QA/QC filter

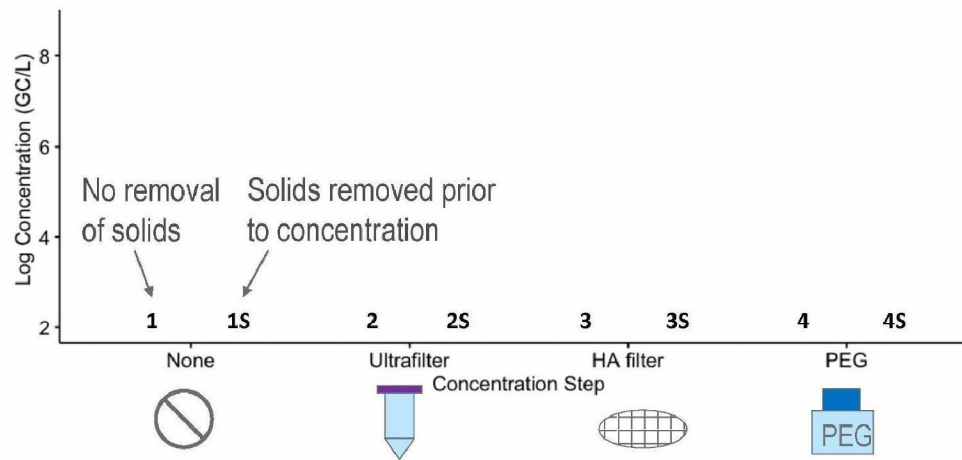
With Recovery Correction



- Conclusions:
  - Across all groups, 80% of the values fall within +/- 1-log range

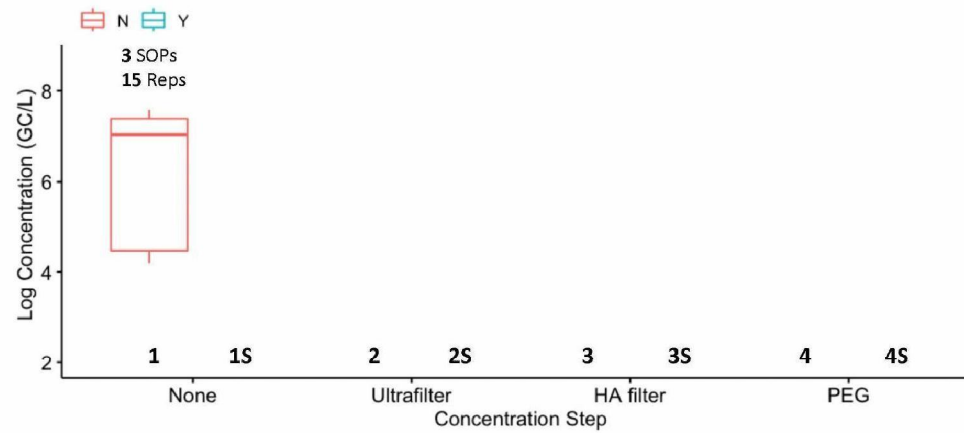
## Reproducibility within a method group

- Eight method groups based on solids removal and concentration step



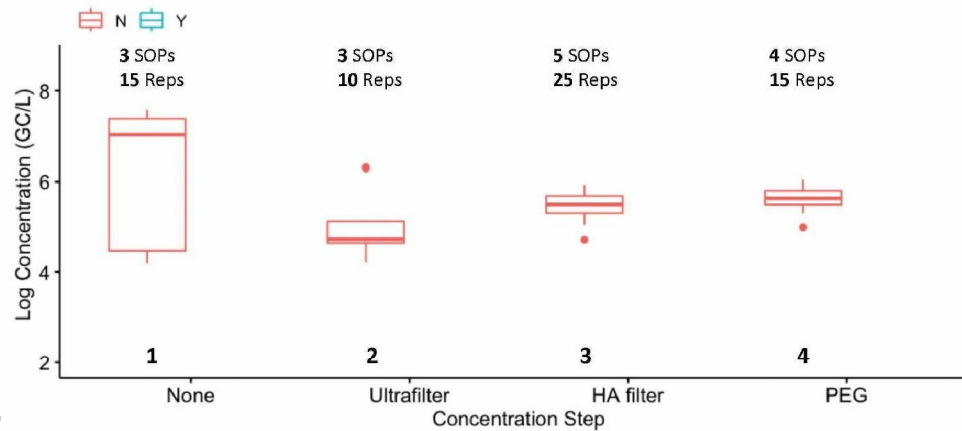
## Reproducibility within a method group

- Eight method groups based on solids removal and concentration step



## Reproducibility within a method group

- Eight method groups based on solids removal and concentration step

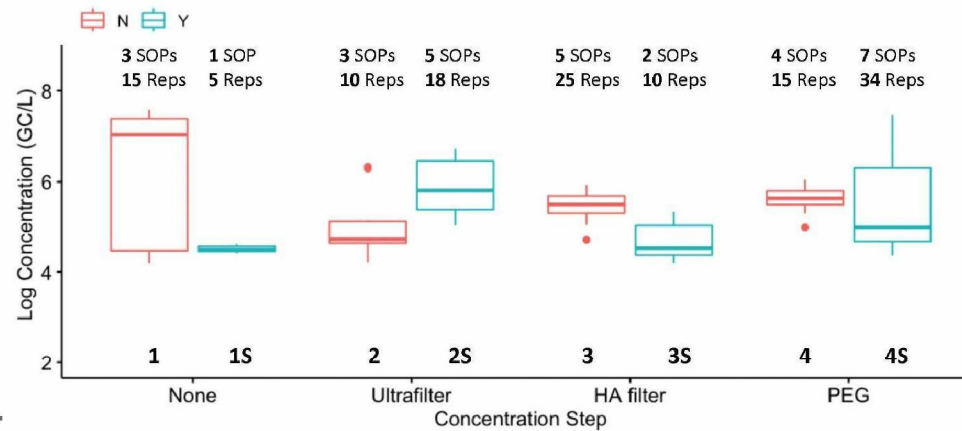


- Conclusions:

- Correcting for recovery generally brings the concentration methods in line with no-concentration methods

## Reproducibility within a method group

- Eight method groups based on solids removal and concentration step



- Conclusions:

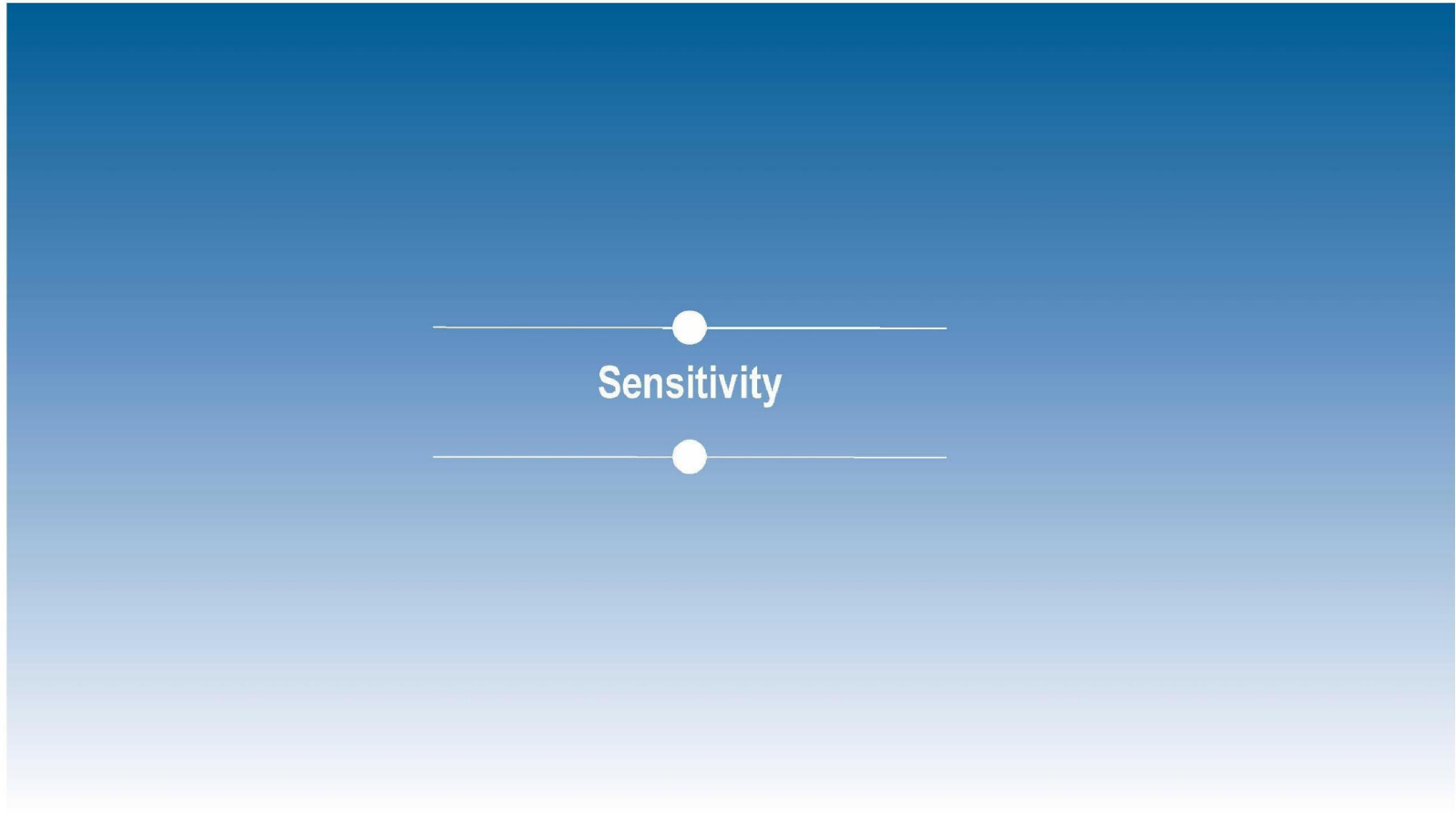
- Correcting for recovery generally brings the concentration methods in line with no-concentration methods
  - No systematic impact from solids removal step

## Reproducibility within a SOP

- Precision evaluated based on variability in replicates run for each method

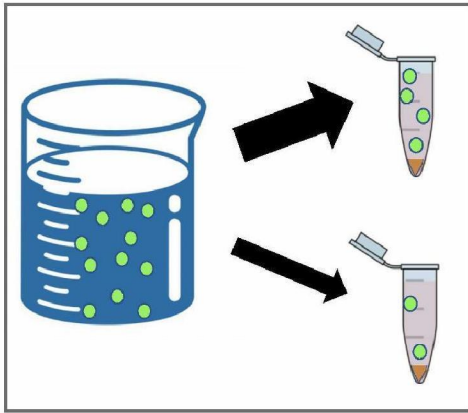
SARS-CoV-2 Target	Standard deviation of replicates (in log GC/L)	
	Uncorrected	Recovery-Corrected
N1	<b>0.15</b> [0.04 – 0.38]	<b>0.13</b> [0.032 – 0.60]
N2	<b>0.14</b> [0.01 – 0.53]	<b>0.13</b> [0.033 – 0.51]

- Conclusions:
  - *Precision within a lab is high based on ~5 replicates*
  - *Higher precision makes it easier to identify differences in raw wastewater concentrations over time*

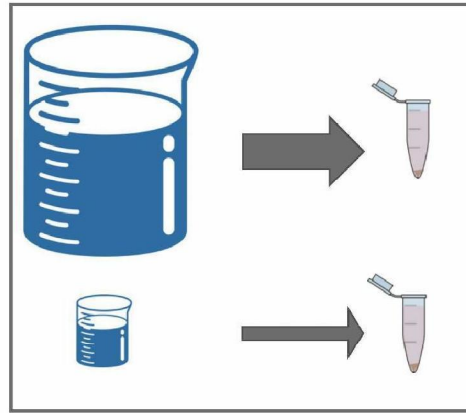


# Method Sensitivity

Recovery Efficiency



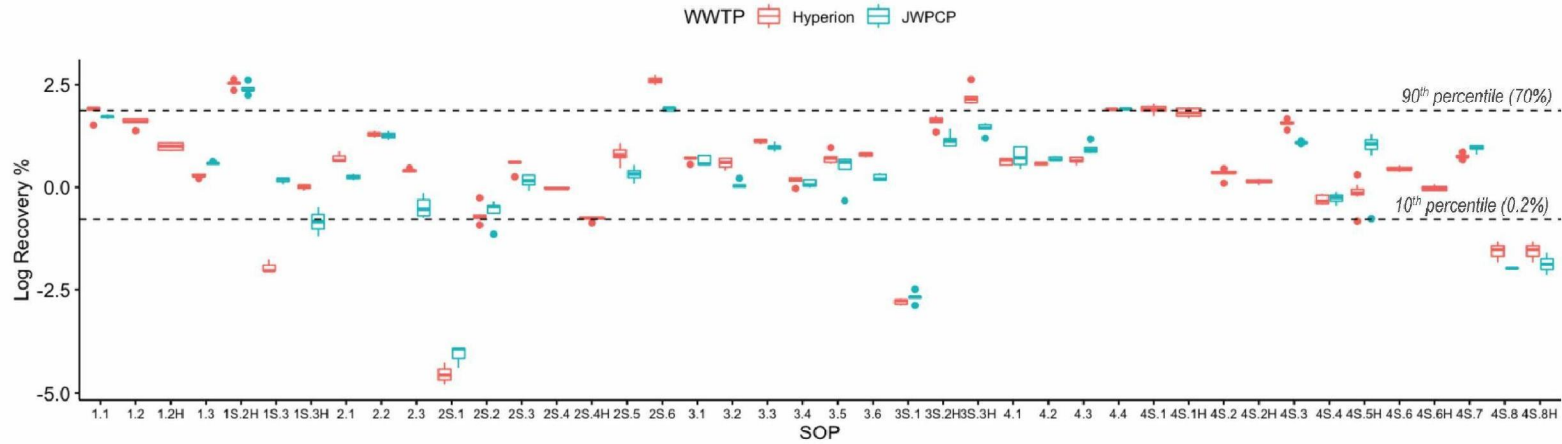
Concentration Factor



Instrument Sensitivity

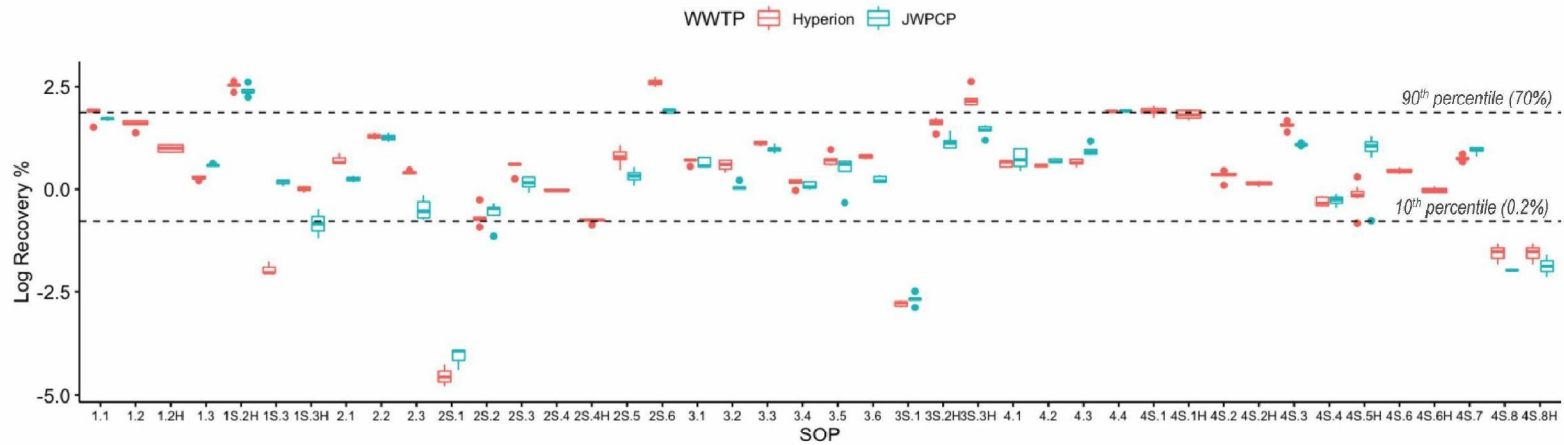


# Recovery Efficiencies



- Conclusions:
  - *Recovery efficiency between two plants was not statistically different*

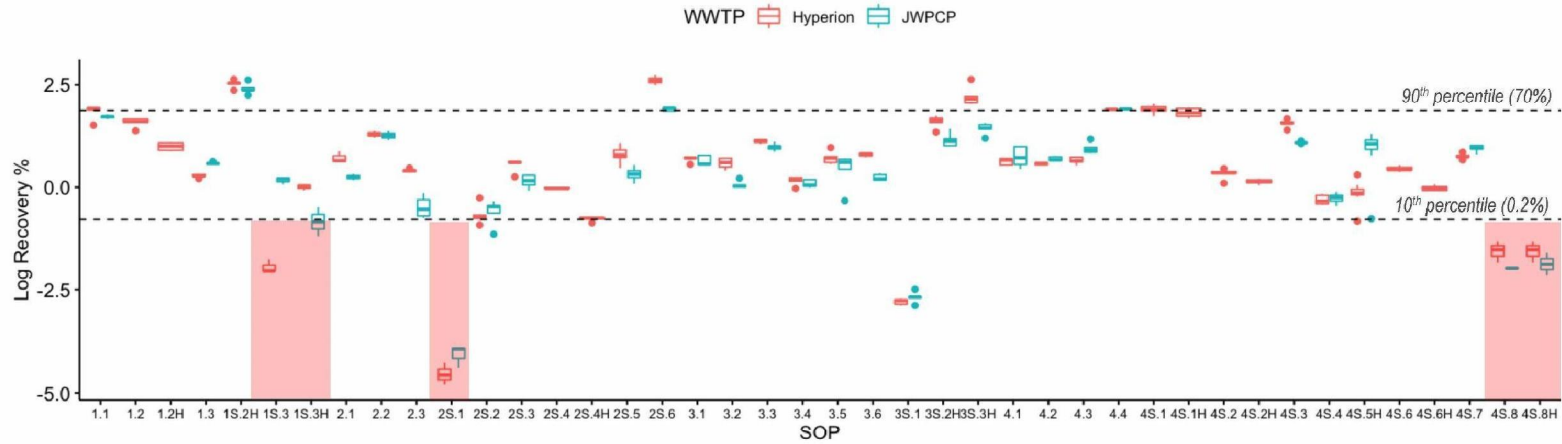
# Recovery Efficiencies



## Conclusions:

- *Recovery efficiency between two plants was not statistically different*
- **Methods show a wide range of recovery efficiencies (7 orders of magnitude)**

# Recovery Efficiencies

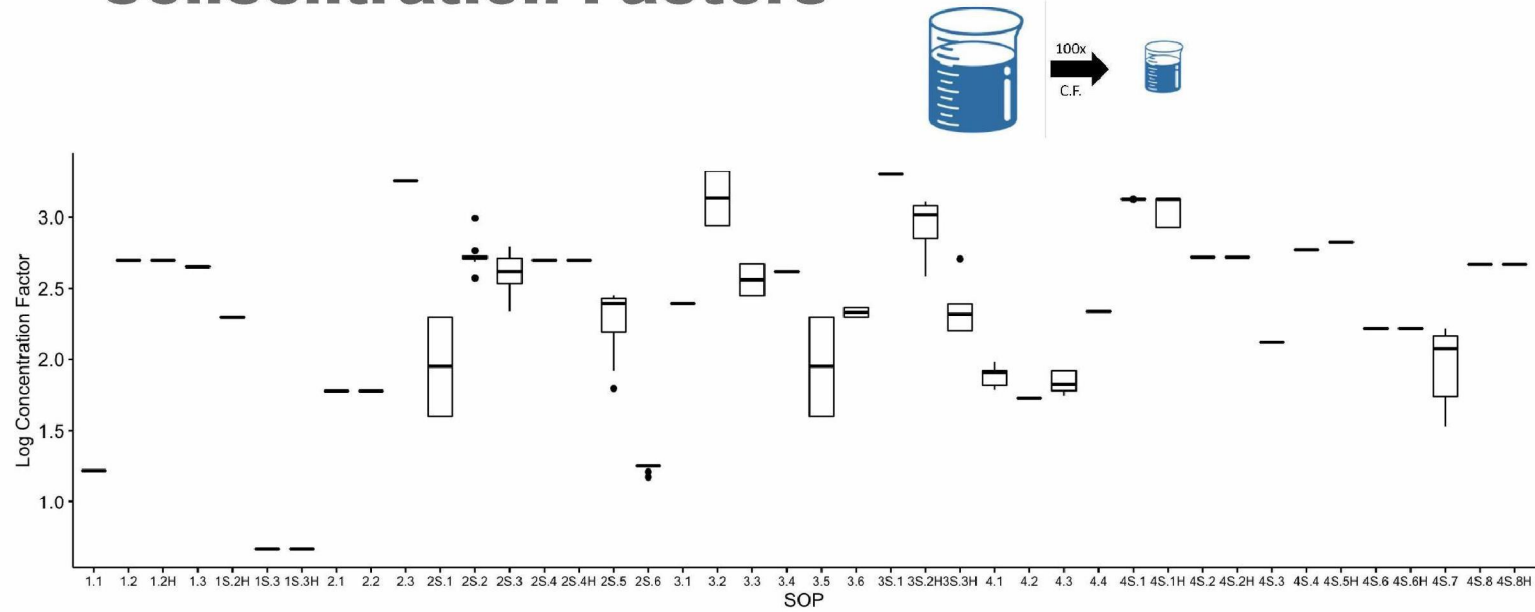


	1.1	1.2	1.2H	1.3	1S.2H	1S.3	1S.3H	2.1	2.2	2.3	2S.1	2S.2	2S.3	2S.4	2S.4H	2S.5	2S.6	3.1	3.2	3.3	3.4	3.5	3.6	3S.1	3S.2H	3S.3H	4.1	4.2	4.3	4.4	4S.1	4S.1H	4S.2	4S.2H	4S.3	4S.4	4S.5H	4S.6	4S.6H	4S.7	4S.8	4S.8H		
Hyperion NDs		1/10			6/6	6/6	1/6	1/6			10/10	4/10	1/10		1/10	2/10	7/10				1/10								4/10									1/10	1/10	11/16			6/6	6/6
JWPCP NDs	X	X			2/10	6/6	6/6	4/4	2/6		10/10	10/10	3/10	X	X	7/10	10/10				1/10	6/10		2/10		3/10	1/10		5/10		X	X	X	X	10/10	4/10	6/20	X	X	7/10	6/6	6/6		

■ Conclusions:

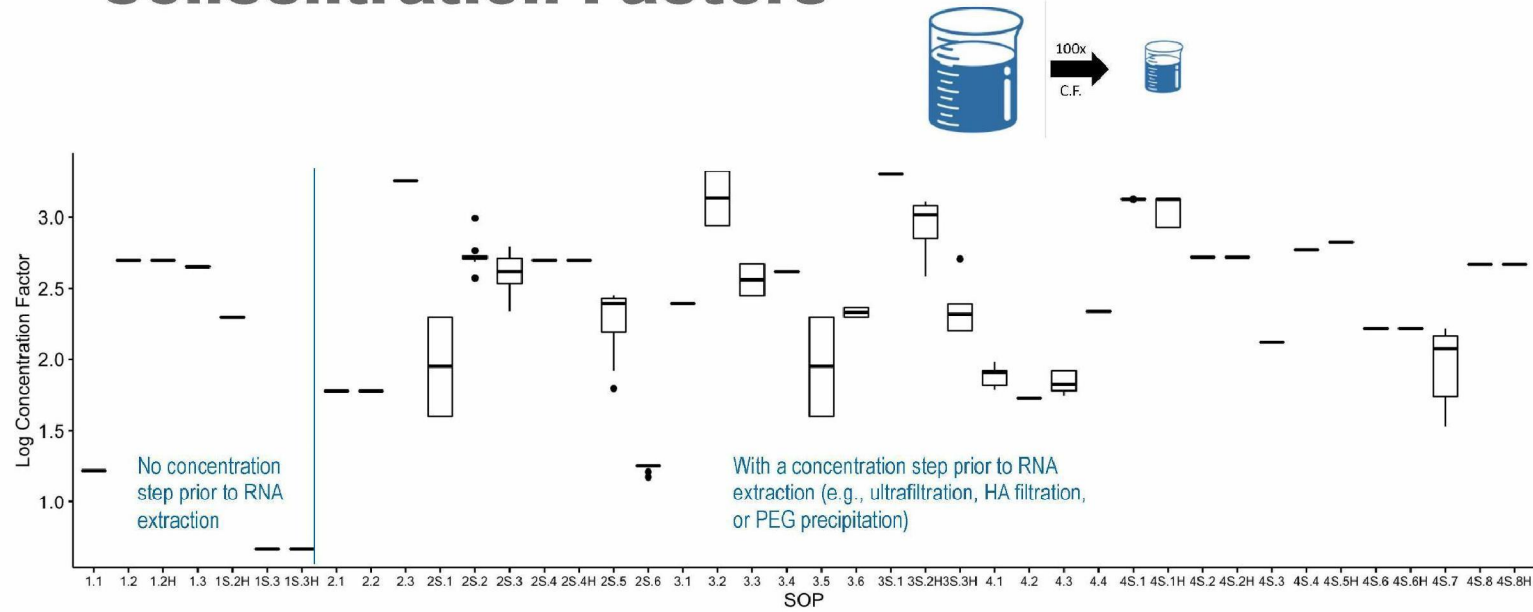
- Recovery efficiency between two plants was not statistically different
- Methods show a wide range of recovery efficiencies (7 orders of magnitude)
- Methods with lower recovery efficiencies more likely to produce non-detects (NDs)

# Concentration Factors



- Conclusions:
  - *Methods show a wide range of concentration factors (>2 orders of magnitude)*

# Concentration Factors

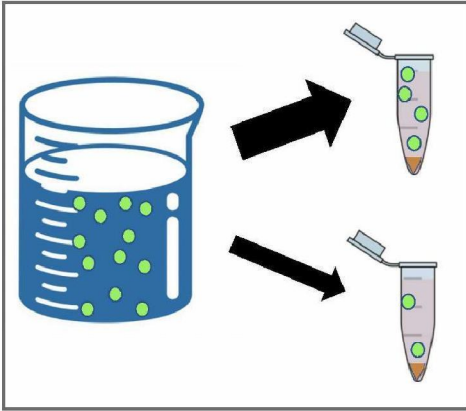


■ **Conclusions:**

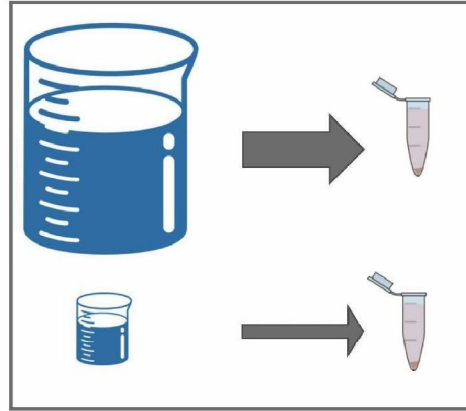
- *Methods show a wide range of concentration factors (>2 orders of magnitude)*
- *Methods without a concentration step prior to RNA extraction did not always have a lower CF*

## Theoretical Limit of Detection for Methods

Recovery Efficiency



Concentration Factor

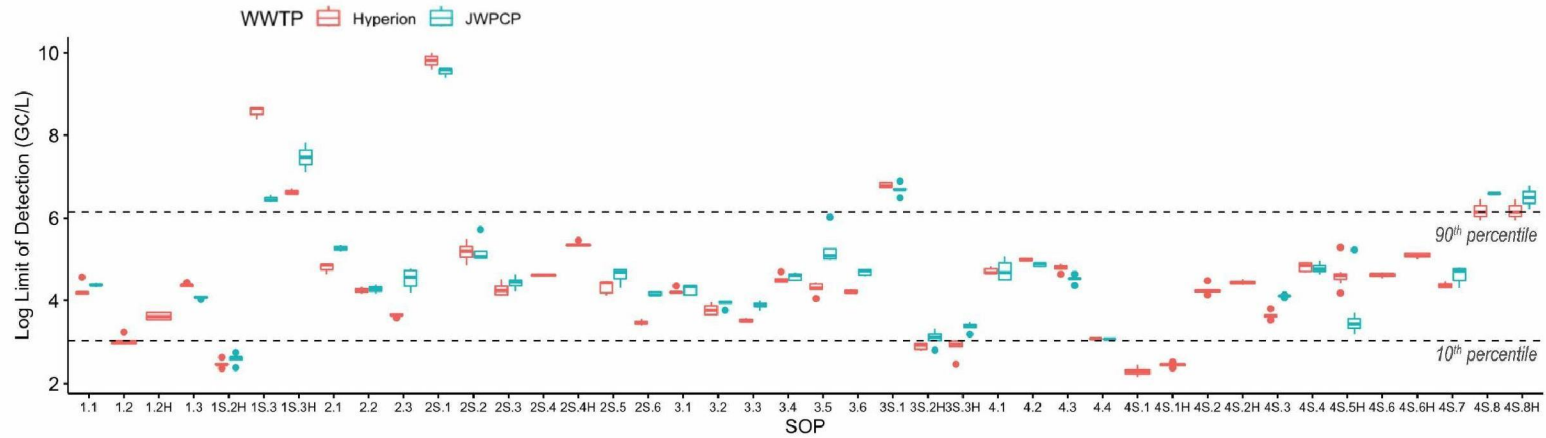


Instrument Sensitivity



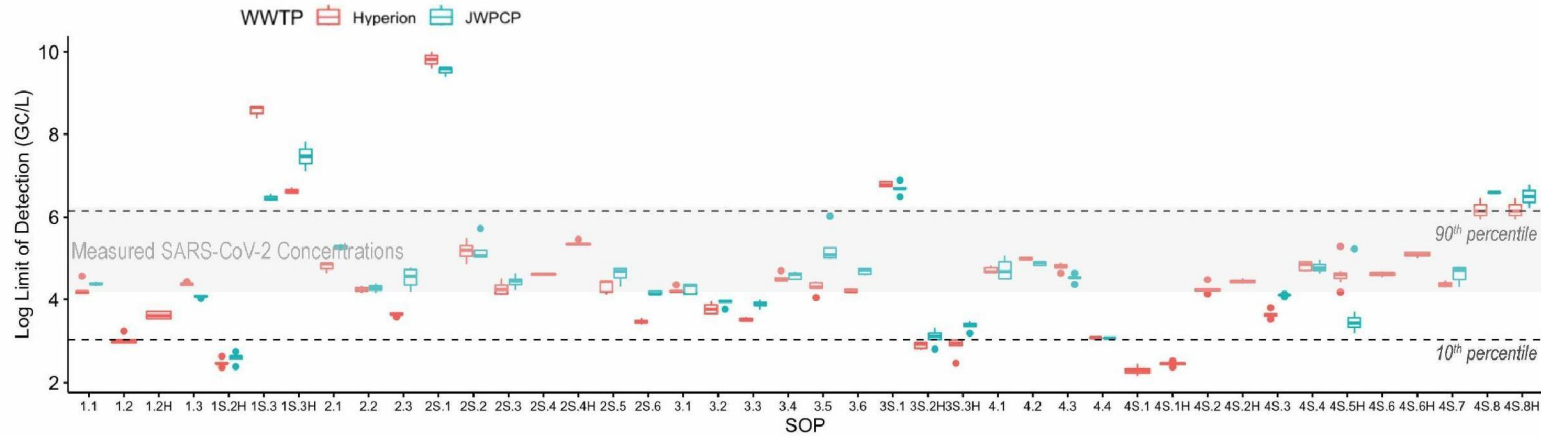
$$\text{Theoretical LOD} = \frac{\text{Instrument Detection Limit (assumed } 1 \frac{\text{GC}}{\text{assay}} \times \frac{\text{assay}}{5\mu\text{L}})}{\text{Concentration Factor} \times \text{Recovery}}$$

## Theoretical Limit of Detection for Methods



- Conclusions:
  - *Limit of detection spanned 7-orders of magnitude*

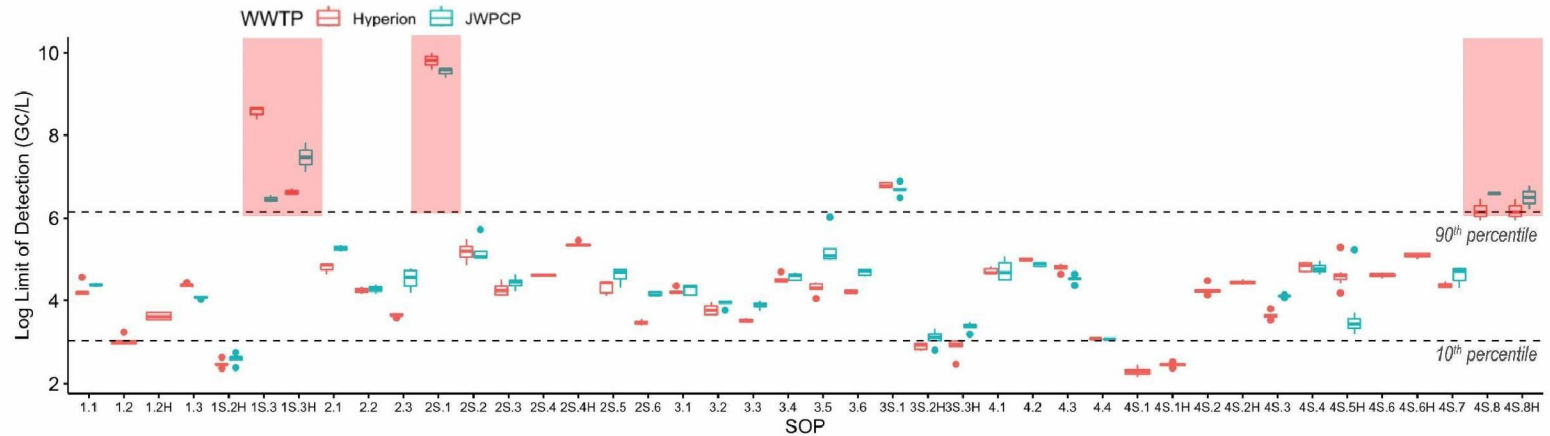
# Theoretical Limit of Detection for Methods



	1.1	1.2	1.2H	1.3	1S.2H	1S.3	1S.3H	2.1	2.2	2.3	2S.1	2S.2	2S.3	2S.4	2S.4H	2S.5	2S.6	3.1	3.2	3.3	3.4	3.5	3.6	3S.1	3S.2H	3S.3H	4.1	4.2	4.3	4.4	4S.1	4S.1H	4S.2	4S.2H	4S.3	4S.4	4S.5H	4S.6	4S.6H	4S.7	4S.8	4S.8H		
Hyperion NDs		1/10				6/6	6/6	1/6	1/6		10/10	4/10	1/10		1/10	2/10	7/10				1/10								4/10								1/10	1/10	11/16				6/6	6/6
JWPCP NDs		X	X		2/10	6/6	6/6	4/4	2/6		10/10	10/10	3/10	X	X	7/10	10/10			1/10	6/10		2/10		3/10			1/10		5/10		X	X	X	X	10/10	4/10	6/20	X	X	7/10	6/6	6/6	

- Conclusions:
  - *Limit of detection spanned 7-orders of magnitude*
  - *Methods generally able to quantify a 10-fold lower concentration than those in August, 2020*

# Theoretical Limit of Detection for Methods

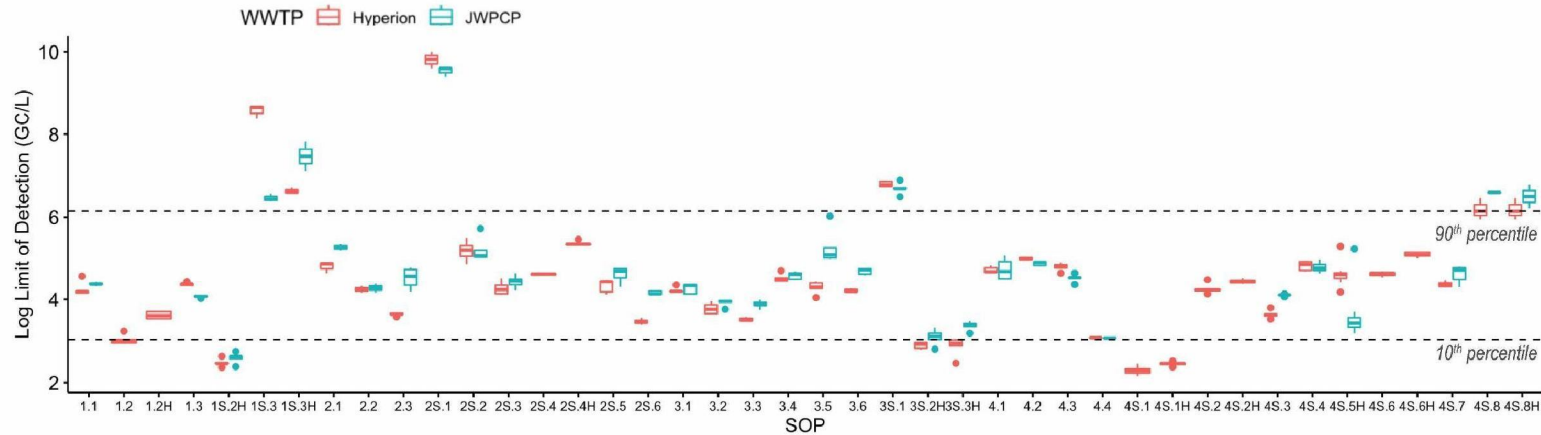


	1.1	1.2	1.2H	1.3	1S.2H	1S.3	1S.3H	2.1	2.2	2.3	2S.1	2S.2	2S.3	2S.4	2S.4H	2S.5	2S.6	3.1	3.2	3.3	3.4	3.5	3.6	3S.1	3S.2H	3S.3H	4.1	4.2	4.3	4.4	4S.1	4S.1H	4S.2	4S.2H	4S.3	4S.4	4S.5H	4S.6	4S.6H	4S.7	4S.8	4S.8H			
Hyperion NDs		1/10				6/6	6/6	1/6	1/6		10/10	4/10	1/10		1/10	2/10	7/10				1/10								4/10									1/10	1/10	11/16				6/6	6/6
JWPCP NDs		X	X		2/10	6/6	6/6	4/4	2/6		10/10	10/10	3/10	X	X	7/10	10/10				1/10	6/10		2/10		3/10		1/10		5/10		X	X	X	X		10/10	4/10	6/20	X	X	7/10	6/6	6/6	

■ Conclusions:

- *Limit of detection spanned 7-orders of magnitude*
- *Methods generally able to quantify a 10-fold lower concentration than those in August, 2020*
- *Methods with all NDs had high LODs (lower sensitivity)*

# Theoretical Limit of Detection for Methods

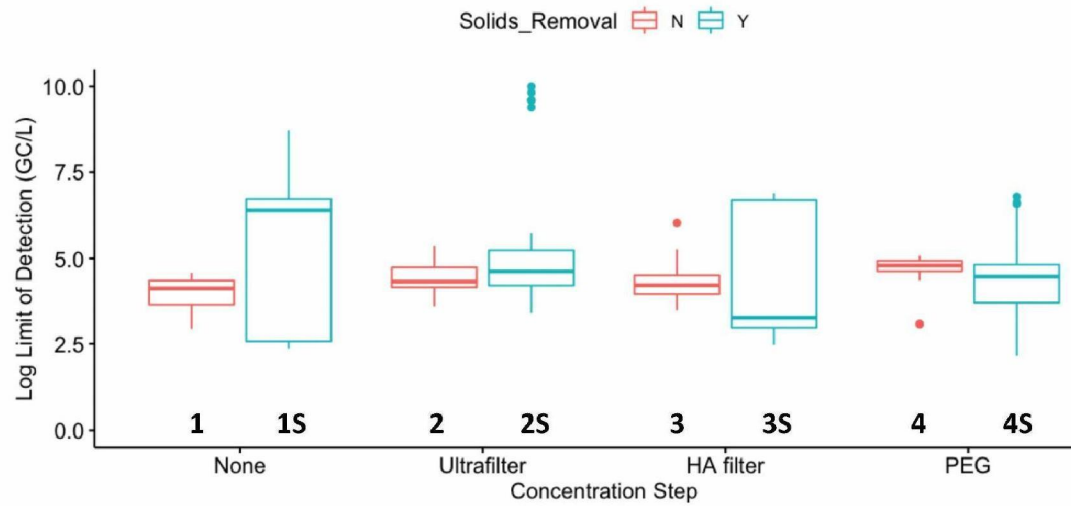


	1.1	1.2	1.2H	1.3	1S.2H	1S.3	1S.3H	2.1	2.2	2.3	2S.1	2S.2	2S.3	2S.4	2S.4H	2S.5	2S.6	3.1	3.2	3.3	3.4	3.5	3.6	3S.1	3S.2H	3S.3H	4.1	4.2	4.3	4.4	4S.1	4S.1H	4S.2	4S.2H	4S.3	4S.4	4S.5H	4S.6	4S.6H	4S.7	4S.8	4S.8H		
Hyperion NDs		1/10				6/6	6/6	1/6	1/6		10/10	4/10	1/10		1/10	2/10	7/10				1/10								4/10								1/10	1/10	11/16				6/6	6/6
JWPCP NDs		X	X		2/10	6/6	6/6	4/4	2/6		10/10	10/10	3/10	X	X	7/10	10/10				1/10	6/10		2/10		3/10		1/10		5/10		X	X	X	X	10/10	4/10	6/20	X	X	7/10	6/6	6/6	

■ Conclusions:

- *Limit of detection spanned 7-orders of magnitude*
- *Methods generally able to quantify a 10-fold lower concentration than those in August, 2020*
- *Methods with all NDs had high LODs (lower sensitivity)*
- **Low LOD is important for tracking trends over a range of concentrations**

## Theoretical Limit of Detection by Method Group

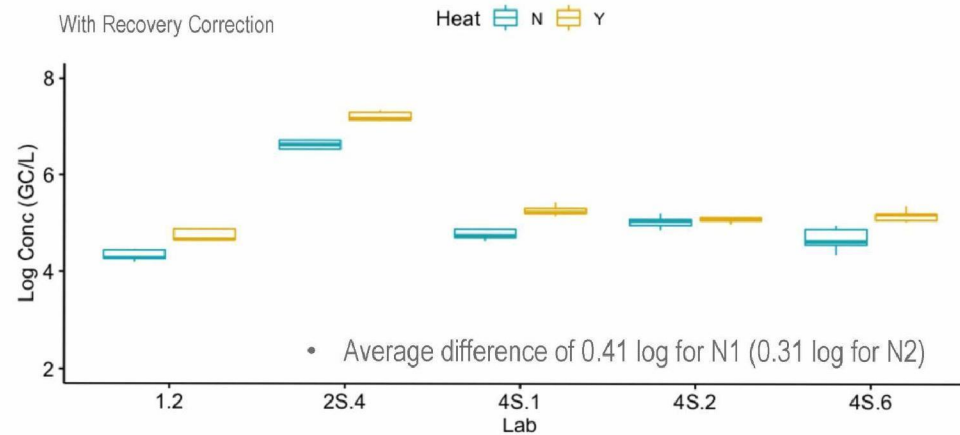


- Conclusions:
  - *The SOPs with highest sensitivity were not all associated with the same method group*
  - *Multiple methods may be capable of achieving high sensitivities*



Impact of Other Method Steps

# Impact of Pasteurization



## Conclusions:

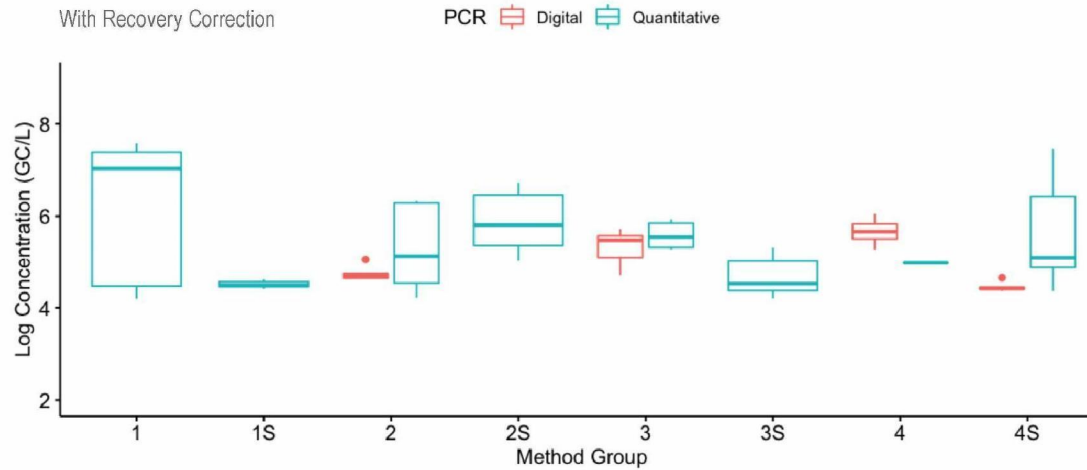
- *Pasteurization at 60°C for 60 minutes led to a significant but small increase in SARS-CoV-2 number*
- *Concern that pasteurization would degrade the signal – results show no clear impact on sensitivity*
- *Importance: ability to pasteurize may open the door to more labs being able to test for SARS-CoV-2*

## Impact of Primer Set

Round	Significant difference between N1 and N2?	Log difference (N1- N2)
Plant 1	Yes (p = 1e-8)	0.13
Plant 2	Yes (p = 0.00042)	0.12

- Conclusions:
  - *While significant, the impact of selecting primer set N1 or N2 is small compared to other sources of variability*
  - *Importance: may not be necessary to run both primer sets when quantifying SARS-CoV-2 concentrations*

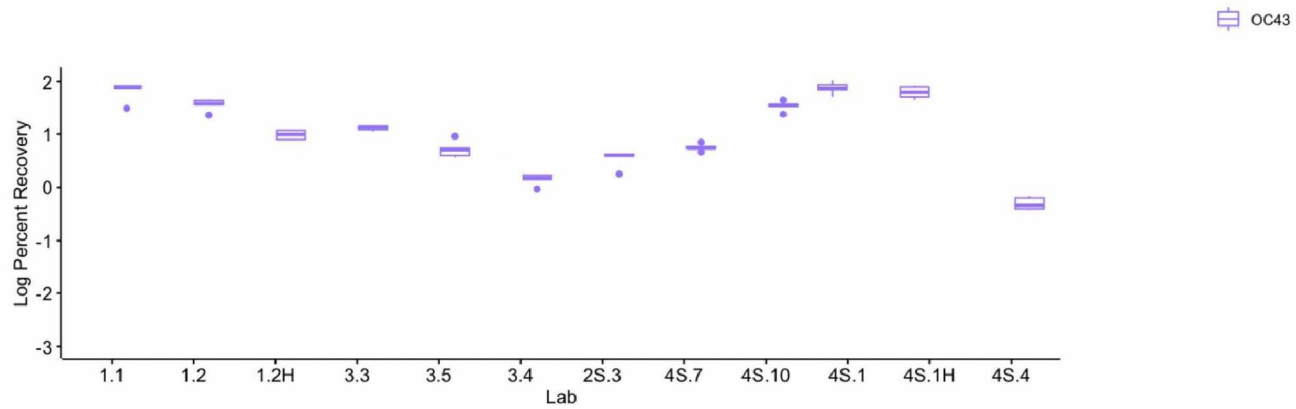
## Impact of PCR Platform



- Conclusions:
  - *No clear patterns emerged between the two quantification platforms*
  - *Merits further research to evaluate impacts on inhibition and sensitivity*

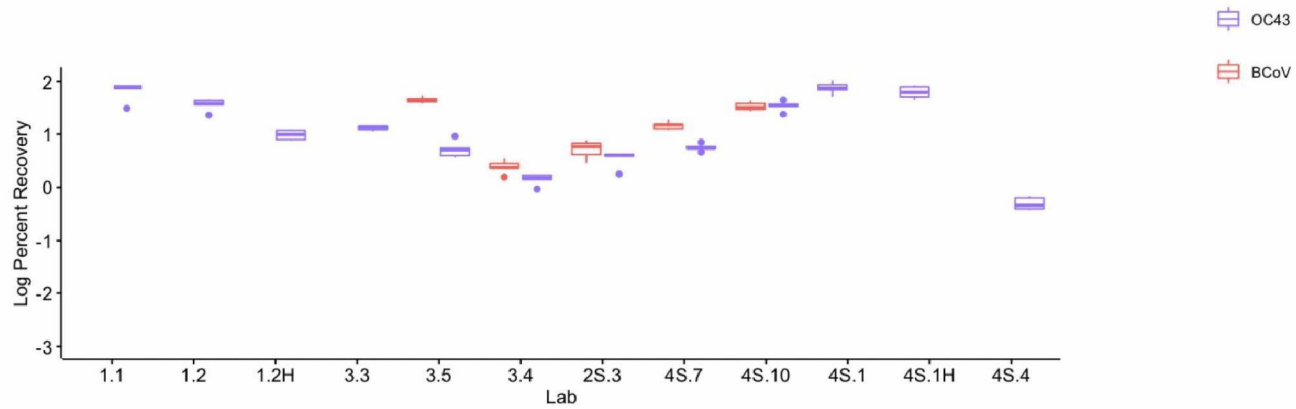
## Impact of Matrix Spike Selection

- Evaluated impact of matrix spike surrogate on SARS-CoV-2 findings



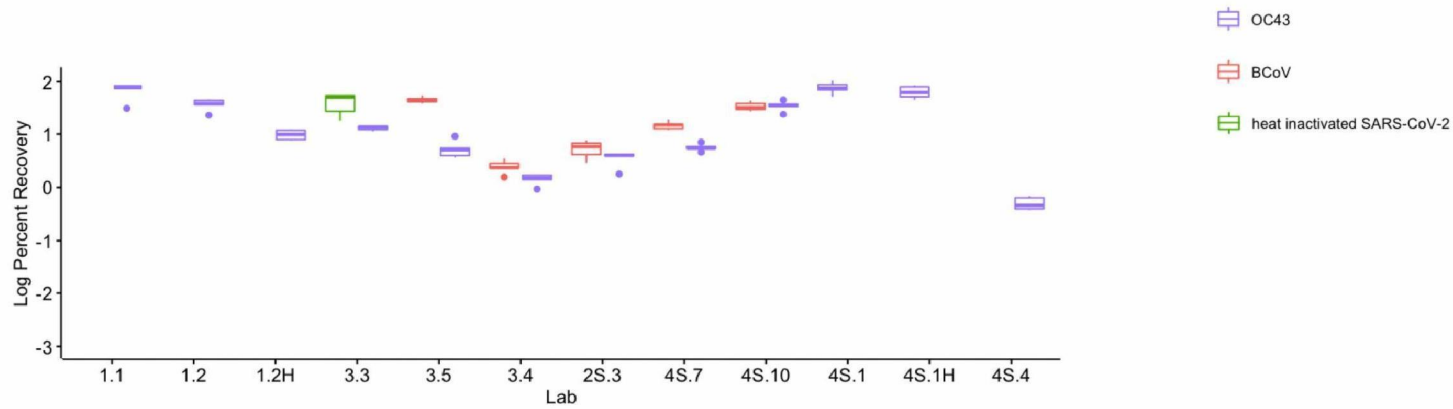
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## Impact of Matrix Spike Selection

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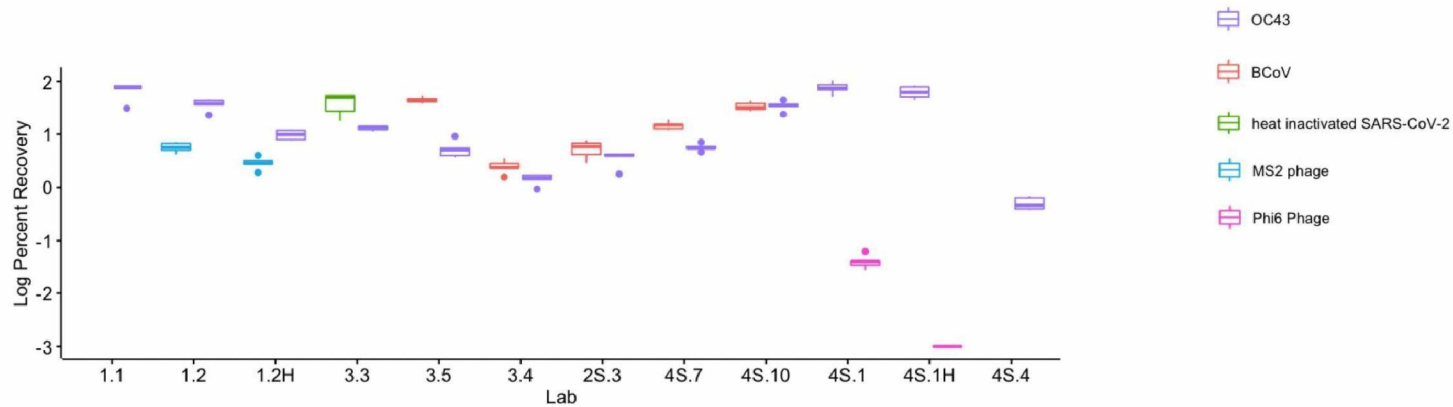


- Conclusions:

- OC43 showed similar behavior to other betacoronaviruses (bovine coronavirus and heat-inactivated SARS-CoV-2)

# Impact of Matrix Spike Selection

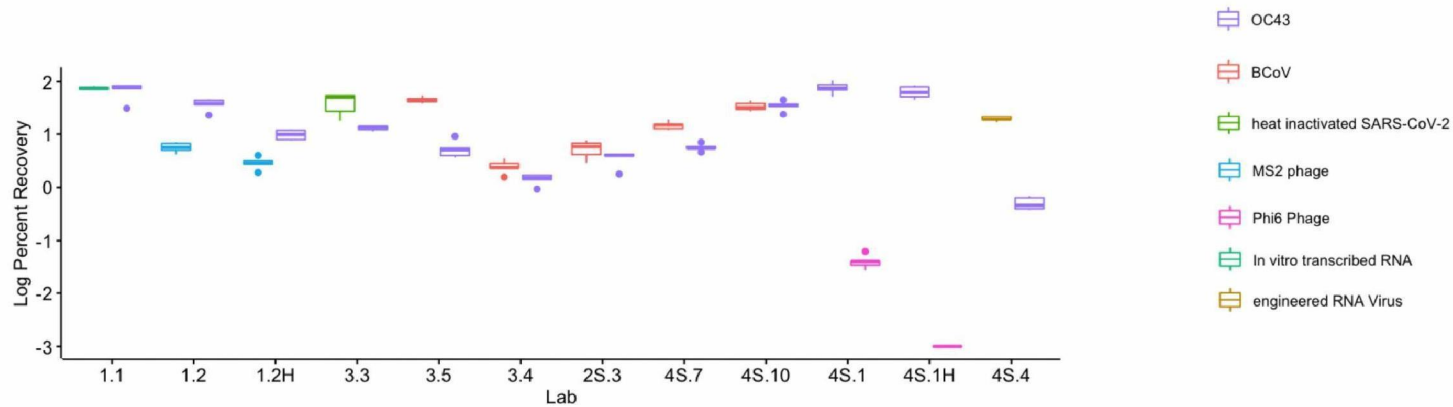
- Evaluated impact of matrix spike surrogate on SARS-CoV-2 findings



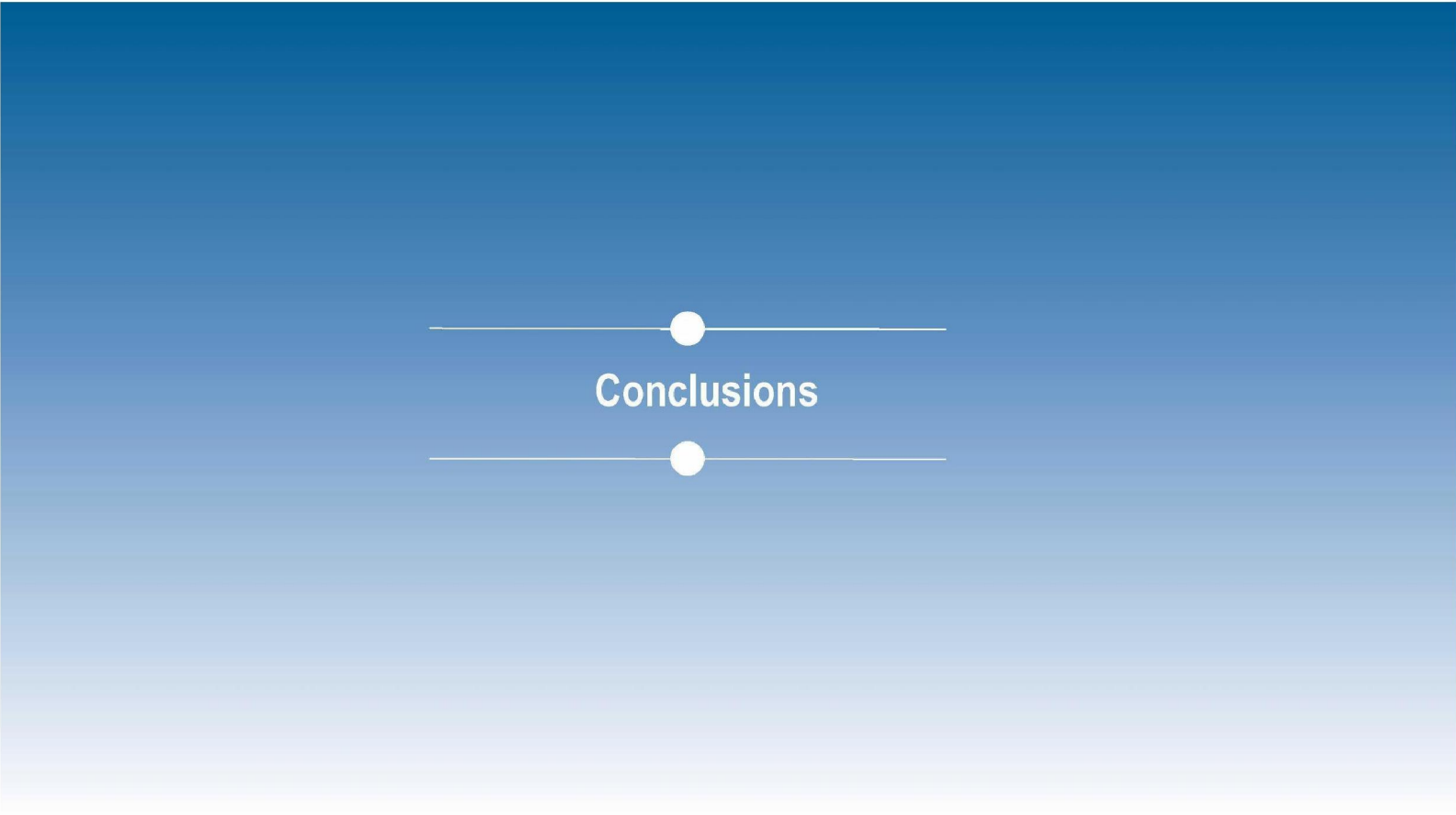
- Conclusions:
  - OC43 showed similar behavior to other betacoronaviruses (bovine coronavirus and heat-inactivated SARS-CoV-2)
  - OC43 showed greater similarity to MS2 bacteriophage than Phi6 in the methods tested

# Impact of Matrix Spike Selection

- Evaluated impact of matrix spike surrogate on SARS-CoV-2 findings



- Conclusions:
  - OC43 showed similar behavior to other betacoronaviruses (bovine coronavirus and heat-inactivated SARS-CoV-2)
  - OC43 showed greater similarity to MS2 bacteriophage than Phi6 in the methods tested
  - Multiple surrogates may be acceptable, but additional work needed to understand similarities with SARS-CoV-2



Conclusions

## Conclusions

- Nationwide interlaboratory method comparison showed high reproducibility
  - *Multiple methods may be used to obtain reproducible results*
  - *The same SOP or lab should be used to track trends at a given location*
- Quality assurance plans are critical for reproducibility
  - *Recovery efficiencies varied by 7 orders of magnitude*
  - *Matrix spikes critical to quantify recovery and obtain reproducible numbers*
- Study showed no systematic impact from key differences between methods
  - *Minimal impact of solids removal, concentration, pasteurization, primer selection*
- Findings support use of wastewater surveillance for tracking trends
  - *Methods with higher sensitivity allow tracking over a wider range of concentrations*

## Next Steps

- Sites will have different requirements and constraints during selection of methods
- Additional criteria should be used to select the “best” method for your application
  - *Sensitivity*
  - *Cost*
  - *Operator experience*
  - *Material requirements*
  - *Throughput or processing time*
- Address other knowledge gaps for wastewater-based epidemiology
- Continued coordination on methods is encouraged

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- Trussell Technologies

- [Redacted] 5.1.2e
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- [Redacted] 5.1.2e

- BCS Labs

- [Redacted] 5.1.2e
- [Redacted] 5.1.2e

- Cel Analytical

- [Redacted] 5.1.2e
- [Redacted] 5.1.2e

- WRF Project Advisory Committee

## SARS-CoV-2 Interlaboratory Consortium

- *Biological Consulting Services (BCS) Laboratories*
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- *City University of New York*
- *Columbia University*
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- *IDEXX Laboratories, Inc.*
- *Los Angeles County Sanitation District*
- *Michigan State University*
- *Mycometrics*
- *New York City Department of Environmental Protection*
- *Ohio State University*
- *Oregon State University*
- *Promega Corporation*
- *Saginaw Valley State University*
- *SiREM*
- *Source Molecular Corporation*
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- *Tulane University*
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- *University of Utah*
- *University of Wisconsin*
- *Utah State University*
- *Weck Labs*
- *Wisconsin State Lab of Hygiene*

# Thank You

**See publications for additional details**

Pre-publication available at medRxiv:

<https://www.medrxiv.org/content/10.1101/2020.11.02.20221622v1>

In peer review at *Environmental Science: Water Research & Technology*